

Synthesis of a ^{125}I -labelled Derivative of 25-Hydroxyvitamin D_3

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The facile synthesis of a radioactive 25-hydroxyvitamin D_3 derivative which is labelled with ^{125}I -Bolton-Hunter Reagent is described.

It is known that vitamin D_3 is metabolized to 25-hydroxy vitamin D_3 (25OHD_3) in the liver and then to 1,25-dihydroxy vitamin D_3 [$1,25(\text{OH})_2\text{D}_3$] in the kidney. The compound $1,25(\text{OH})_2\text{D}_3$ is considered to be a hormone-like substance.^{1,2}

Radioisotope labelled analogues of active vitamin D_3 [$1,25(\text{OH})_2\text{D}_3$, 25OHD_3] are found to be very useful endocrinologically. However only ^3H -labelled derivatives are known,³ but these have low radioactivity, and so highly radioactive ^{125}I -labelled derivatives have been required for some time.

Chemical conversion of vitamin D_3 is difficult owing to its extreme lability, therefore syntheses of vitamin D_3 derivatives have been mostly from steroidal compounds with many steps.

Therefore we investigated a facile synthesis of a ^{125}I -labelled derivative under mild conditions with 25OHD_3 (1) as the starting material (Scheme 1). Treatment of fluoren-9-ylmethoxycarbonyl (Fmoc) protected β -alanine with trimethylacetyl chloride in anhydrous tetrahydrofuran (THF) in the presence of dimethylaminopyridine (DMAP)^{4,5} following esterification of 25OHD_3 (1) successfully afforded the derivative (2).† The isolated yield of (2) was 72.2%. The

† Selected spectroscopic data for (2): ^1H n.m.r. (90 MHz) and u.v.: $\delta(\text{CDCl}_3)$ 0.54 (3H, s, CH_3 -18), 3.30–3.60 (2H, m, $-\text{CH}_2-\text{N}-$), 4.10–4.50 (3H, m, Fmoc), 4.80–5.60 (4H, m, H-19E, H-3 α , H-19Z, $-\text{NH}-$), 5.94–6.29 (2H, m, H-7, H-6), 7.20–7.81 (8H, m, Fmoc), λ_{max} (EtOH) nm 300.2, 266.4, 214.4. (3): ^1H n.m.r. (90 MHz) and u.v.: $\delta(\text{CDCl}_3)$ 0.54 (3H, s, CH_3 -18), 2.36–2.52 (2H, m, $-\text{CO}-\text{CH}_2-$), 2.90–3.05 (2H, m, $-\text{CH}_2-\text{N}-$), 3.50–4.00 (2H, b, $-\text{NH}_2$), 4.86–5.08 (3H, m, H-19E, H-3 α , H-19Z), 5.90–6.30 (2H, m, H-7, H-6), λ_{max} (EtOH) nm 264.5. (5): ^1H n.m.r. (400 MHz) and u.v.: $\delta(\text{CHCl}_3)$ 0.52 (3H, s, CH_3 -18), 0.92 (3H, d, CH_3 -21, J 6.3 Hz), 1.23 (6H, s, CH_3 -26, 27) 2.56–2.20 (8H, m, H-4 α , $-\text{NCO}-\text{CH}_2-$, H-1 β , H-4 β , $-\text{OCO}-\text{CH}_2-$, H-1 α), 2.85–2.73 (3H, m, $-\text{CH}_2-\text{Ph}$, H-9 β), 3.48 (2H, m, $-\text{CH}_2-\text{N}-$), 4.86 (1H, d, H-19E, J 2.0 Hz), 5.01 (1H, m, H-3 α), 5.09 (1H, m, H-19Z), 5.94 (1H, t, $-\text{NH}-$, J 5.9 Hz), 6.01 (1H, d, H-7, J 11.2 Hz), 6.20 (1H, d, H-6, J 11.2 Hz), 6.87 (1H, d, H-Ph, J 8.3 Hz), 4.04 (1H, dd, H-Ph, J 2.0 and 8.3 Hz), 7.47 (1H, d, H-Ph, J 2.0 Hz), λ_{max} (EtOH) nm 268.2.

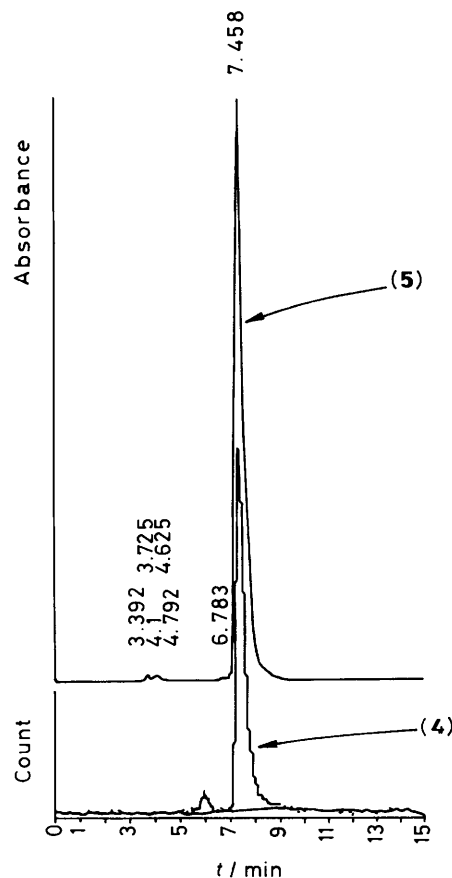
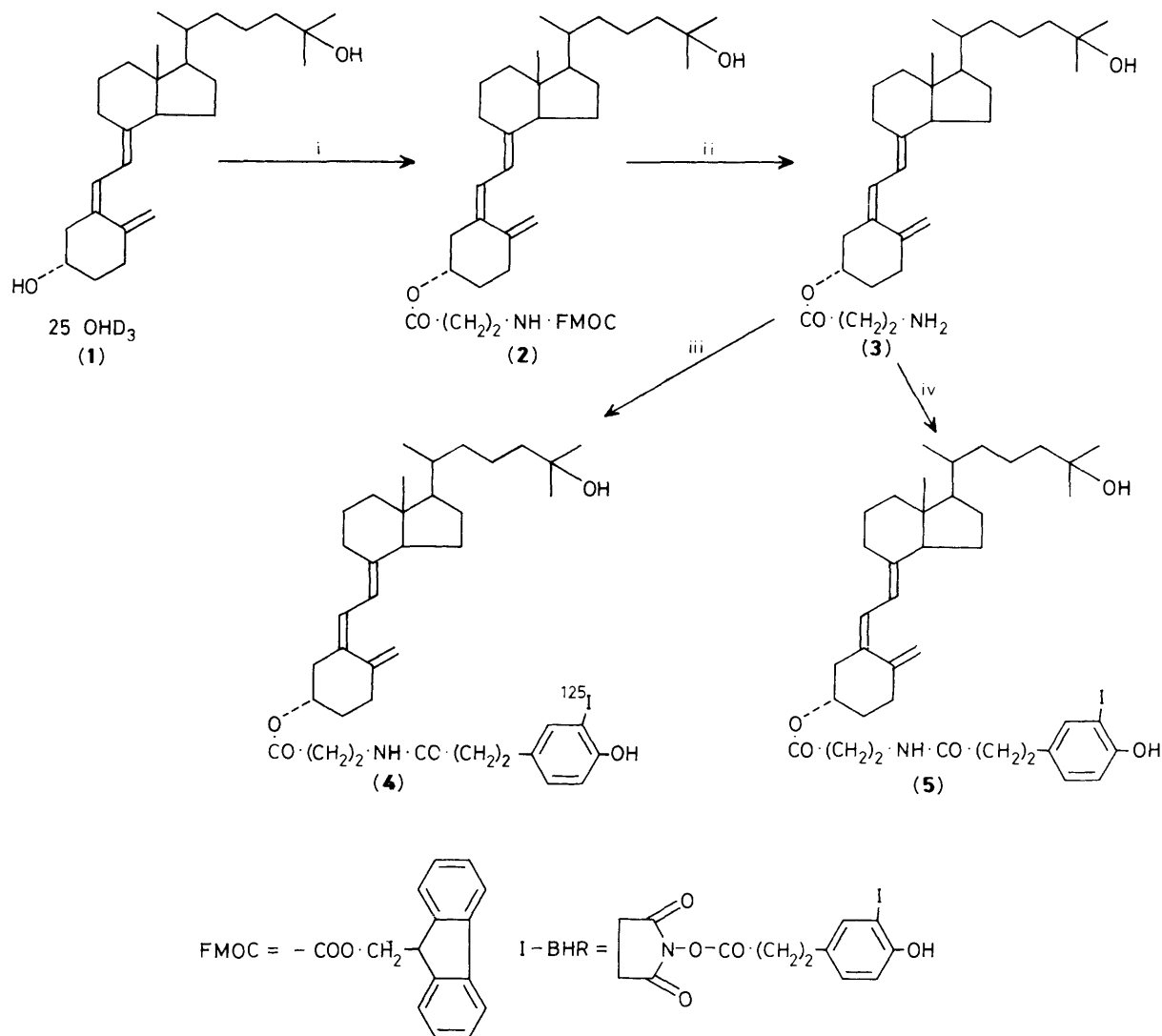


Figure 1. H.p.l.c. of compounds (4) and (5). Detection of (4): (ALOKA) Radioanalyzer RLC-551, (ALOKA) γ -rayflow detector FGD-101. Detection of (5): (SHIMADZU) u.v. spectrophotometric detector SPD-6A, 265 nm. H.p.l.c. conditions: (MISTUITOASTU) Zorbax Pro10 (silica), 4.6×250 mm, (SHIMADZU) Guardcolumn, 4.6×50 mm, 20% isopropanol-*n*-hexane 1 ml/min.



Scheme 1. Reagents and conditions: i, β -Ala-FMOC, trimethylacetylchloride, DMAP, in anhydrous THF, at 5 °C for 1 h, and then at room temp. for 2 h; ii, morpholine, at room temp. for 2 h; iii, ^{125}I -BHR (NEN, NEX-120-10), Et_3N , in anhydrous THF, at 25 °C for 18 h; iv, I-BHR synthesized in our laboratory, Et_3N , in anhydrous THF, at 5 °C for 16 h.

deprotection of the derivative (2) with morpholine gave compound (3),† with an isolated yield of 83.9%.

In the presence of a base, treatment of compound (3) with ^{125}I -Bolton-Hunter Reagent [*N*-succinimidyl 3-[4-hydroxy-3-(^{125}I) iodophenyl] propionate: NEN NEX-120-10 2200Ci/mmol ($1\text{Ci} = 3.7 \times 10^{10}\text{ Bq}$)],⁶ a protein labelling reagent, gave the ^{125}I -labelled derivative (4) with an isolated radioactive yield of 50–60% which was purified by h.p.l.c. The same treatment of (3) with non-radioactive I-Bolton-Hunter Reagent (I-BHR) [*N*-succinimidyl 3-(4-hydroxy-3-iodophenyl)propionate] gave the corresponding compound (5).† The isolated yield, purified by h.p.l.c., of (5) was 47.2%. Compounds (4) and (5) had an identical retention time for h.p.l.c. (Figure 1).

The ethanol solution of (4) was stable for 60 days at -20°C . The radioactivity of (4) can be calculated at 2200Ci/mmol as the radioactivity of the ^{125}I -BHR is known. This radioactivity of (4) is ten times greater than that of ^3H -labelled 25OHD₃ which has been used previously as a tracer. These results

indicate that it is possible to use ^{125}I -labelled compound (4) as a tracer with high sensitivity in radiobioassay.

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References

- H. F. DeLuca, *Nutr. Rev.*, 1979, **37**, 161.
- N. Ikekawa, *Med. Chem. Rev.*, 1987, **7**, 333.
- P. F. Neville and H. F. DeLuca, *Biochemistry*, 1966, **5**, 2201; H. F. DeLuca, M. Weller, J. W. Blunt, and P. F. Neville, *Arch. Biochem. Biophys.*, 1968, **124**, 122; T. Suda, H. F. DeLuca, and R. B. Hallick, *Anal. Biochem.*, 1971, **43**, 130; S. Yamada, H. K. Schnoes, and H. F. DeLuca, *ibid.*, 1978, **85**, 34.
- R. Ray, S. A. Holick, and M. F. Holick, *J. Chem. Soc., Chem. Commun.*, 1985, 702; R. Ray, S. Rose, S. A. Holick, and M. F. Holick, *Biochem. Biophys. Res. Commun.*, 1985, **132**, 198; R. Ray, S. A. Holick, N. Hanafin, and M. F. Holick, *Biochemistry*, 1986, **25**, 4729; R. Ray and M. F. Holick, *Steroids*, 1988, **51**, 623.
- A. Hassner, L. R. Krepshi, and V. Alexanian, *Tetrahedron*, 1978, **34**, 2069.
- A. E. Bolton and W. M. Hunter, *Biochem. J.*, 1973, **133**, 529.